advanced bony metastases of prostate cancer owing to their selective deposition at the tumor–bone matrix interface, thereby potentially affecting the function of both tumor and stromal cells in the progressive growth of the metastatic deposit.

RESISTANCE TO CANCER TREATMENTS

Resistance mechanisms to the conventional cytotoxic agents were initially characterized in the late twentieth century as defects in drug uptake, metabolism, or export by tumor cells. The *multidrug resistance* (*mdr*) gene defined in vitro in cell lines exposed to increasing concentrations of drugs led to the definition of a family of transport proteins that, when overexpressed, result in the facile transport of a variety of hydrophobic drugs out of the cancer cell. Although efforts to manipulate this transporter to promote drug residence in tumor cells have been pursued, none are clinically useful at this time. Drug-metabolizing enzymes such as cytidine deaminase are upregulated in resistant tumor cells, and this is the basis for so-called "high-dose cytarabine" regimens in the treatment of leukemia. Another resistance mechanism defined during this era involved increased expression of a drug's target, exemplified by amplification of the dihydrofolate reductase gene, in patients who had lost responsiveness to methotrexate, or mutation of topoisomerase II in tumors that relapsed after topoisomerase II modulator treatment.

A second class of resistance mechanisms involves loss of the cellular apoptotic mechanism activated after the engagement of a drug's target by the drug. This occurs in a way that is heavily influenced by the biology of the particular tumor type. For example, decreased alkylguanine alkyltransferase defines a subset of glioblastoma patients with the prospect of greatest benefit from treatment with temozolomide, but has no predictive value for benefit from temozolomide in epithelial neoplasms. Likewise, ovarian cancers resistant to platinating agents have decreased expression of the proapoptotic gene *bax*. These types of findings have prompted the idea that responsive tumors to chemotherapeutic agents are populated by cells that express drug-related cell death controlling genes, creating in effect a state of "synthetic lethality" of the drug **(Chap. 102e)** with the genes expressed in responsive tumors, analogous to the existence in yeast of mutations that are well tolerated in the absence of a physiologic stressor but become lethal in the presence of that stressor. In the case of tumors, the chemotherapy inducing the cell death response is the analogous physiologic stressor.

A third class of resistance mechanisms emerged from sequencing of the targets of agents directed at oncogenic kinases. Thus, patients with CML resistant to imatinib have acquired mutations in the ATP binding domain of $p210^{bcr\text{-}abl}$ in some cases, leading to the screening and design of agents with activity against the mutant proteins. Entirely analogous resistance mechanisms have emerged in patients with lung cancer treated with the EGFR antagonists gefitinib and erlotinib.

A final category of tumor resistance mechanisms to targeted agents includes the upregulation of alternate means of activating the pathway targeted by the agent. Thus melanomas initially responsive to *BRAF* V600E antagonists such as vemurafenib may reactivate raf signaling by upregulating isoforms that can bypass the variant blocked by the drug. Likewise, inhibition of HER2/neu signaling in breast cancer cells can lead to the emergence of variants with distinct oncogenic signaling pathways such as PI3 kinase. Analogously in NSCLC, EGFR inhibitor treatment leads to the emergence of cells with a predominance of c-met protooncogene–dependent signaling in the resistant tumors.

The susceptibility of a tumor to different treatments as a function of its expression of potential drug targets or their mutational profile has led to efforts to define the dominant pathways driving a patient's tumor by genomic techniques including whole exome sequencing. The difficulty with applying such data to patient treatment is recognizing that these pathways may change during the natural history of a tumor and that different sites in a single patient may have tumors with different patterns of gene mutation.

SUPPORTIVE CARE DURING CANCER TREATMENT

MYELOSUPPRESSION

The common cytotoxic chemotherapeutic agents almost invariably affect bone marrow function. Titration of this effect determines the MTD of the agent on a given schedule. The normal kinetics of blood cell turnover influences the sequence and sensitivity of each of the formed elements. Polymorphonuclear leukocytes (PMNs; $t_{1/2}$ = 6–8 h), platelets ($t_{1/2}$ = 5–7 days), and red blood cells (RBCs; $t_{1/2}$ = 120 days) have most, less, and least susceptibility, respectively, to usually administered cytotoxic agents. The nadir count of each cell type in response to classes of agents is characteristic. Maximal neutropenia occurs 6–14 days after conventional doses of anthracyclines, antifolates, and antimetabolites. Alkylating agents differ from each other in the timing of cytopenias. Nitrosoureas, DTIC, and procarbazine can display delayed marrow toxicity, first appearing 6 weeks after dosing.

Complications of myelosuppression result from the predictable sequelae of the missing cells' function. *Febrile neutropenia* refers to the clinical presentation of fever (one temperature ≥38.5°C or three readings ≥38°C but ≤38.5°C per 24 h) in a neutropenic patient with an uncontrolled neoplasm involving the bone marrow or, more usually, in a patient undergoing treatment with cytotoxic agents. Mortality from uncontrolled infection varies inversely with the neutrophil count. If the nadir neutrophil count is >1000/μL, there is little risk; if <500/μL, risk of death is markedly increased. Management of febrile neutropenia has conventionally included empirical coverage with antibiotics for the duration of neutropenia **(Chap. 104)**. Selection of antibiotics is governed by the expected association of infections with certain underlying neoplasms; careful physical examination (with scrutiny of catheter sites, dentition, mucosal surfaces, and perirectal and genital orifices by gentle palpation); chest x-ray; and Gram stain and culture of blood, urine, and sputum (if any) to define a putative site of infection. In the absence of any originating site, a broadly acting β-lactam with anti-*Pseudomonas* activity, such as ceftazidime, is begun empirically. The addition of vancomycin to cover potential cutaneous sites of origin (until these are ruled out or shown to originate from methicillin-sensitive organisms) or metronidazole or imipenem for abdominal or other sites favoring anaerobes reflects modifications tailored to individual patient presentations. The coexistence of pulmonary compromise raises a distinct set of potential pathogens, including *Legionella*, *Pneumocystis*, and fungal agents that may require further diagnostic evaluations, such as bronchoscopy with bronchoalveolar lavage. Febrile neutropenic patients can be stratified broadly into two prognostic groups. The first, with expected short duration of neutropenia and no evidence of hypotension or abdominal or other localizing symptoms, may be expected to do well even with oral regimens, e.g., ciprofloxacin or moxifloxacin, or amoxicillin plus clavulanic acid. A less favorable prognostic group is patients with expected prolonged neutropenia, evidence of sepsis, and end organ compromise, particularly pneumonia. These patients require tailoring of their antibiotic regimen to their underlying presentation, with frequent empirical addition of antifungal agents if fever and neutropenia persists for 7 days without identification of an adequately treated organism or site.

Transfusion of granulocytes has no role in the management of febrile neutropenia, owing to their exceedingly short half-life, mechanical fragility, and clinical syndromes of pulmonary compromise with leukostasis after their use. Instead, colony-stimulating factors (CSFs) are used to augment bone marrow production of PMNs. Early-acting factors such as IL-1, IL-3, and stem cell factor have not been as useful clinically as late-acting, lineage-specific factors such as granulocyte colony-stimulating factor (G-CSF) or GM-CSF, erythropoietin (EPO), thrombopoietin, IL-6, and IL-11. CSFs may easily become overused in oncology practice. The settings in which their use has been proved effective are limited. G-CSF, GM-CSF, EPO, and IL-11 are currently approved for use. The American Society of Clinical Oncology has developed practice guidelines for the use of G-CSF and GM-CSF **(Table 103e-7)**.

Primary prophylaxis (i.e., shortly after completing chemotherapy to reduce the nadir) administers G-CSF to patients receiving cytotoxic