

2092 activates lipoprotein lipase, which is important in clearing chylomicrons from the bloodstream. Patients with diabetes mellitus who have developed ketoacidosis and patients who are on certain medications such as oral contraceptives may also develop high triglyceride levels. Approximately 0.1–2% of cases of acute pancreatitis are drug related. Drugs cause pancreatitis either by a hypersensitivity reaction or by the generation of a toxic metabolite, although in some cases, it is not clear which of these mechanisms is operative (Table 371-1).

Pathologically, acute pancreatitis varies from *interstitial pancreatitis* (pancreas blood supply maintained), which is generally self-limited to *necrotizing pancreatitis* (pancreas blood supply interrupted), in which the extent of necrosis may correlate with the severity of the attack and its systemic complications. Autodigestion is a currently accepted pathogenic theory; according to this theory, pancreatitis results when proteolytic enzymes (e.g., trypsinogen, chymotrypsinogen, proelastase, and lipolytic enzymes such as phospholipase A₂) are activated in the pancreas acinar cell rather than in the intestinal lumen. A number of factors (e.g., endotoxins, exotoxins, viral infections, ischemia, oxidative stress, lysosomal calcium, and direct trauma) are believed to facilitate premature activation of trypsin. Activated proteolytic enzymes, especially trypsin, not only digest pancreatic and peripancreatic tissues but also can activate other enzymes, such as elastase and phospholipase A₂. Spontaneous activation of trypsin also can occur.

ACTIVATION OF PANCREATIC ENZYMES IN THE PATHOGENESIS OF ACUTE PANCREATITIS

Several recent studies have suggested that pancreatitis is a disease that evolves in three phases. The *initial phase* is characterized by intrapancreatic digestive enzyme activation and acinar cell injury. Trypsin activation appears to be mediated by lysosomal hydrolases such as cathepsin B that become colocalized with digestive enzymes in intracellular organelles; it is currently believed that acinar cell injury is the consequence of trypsin activation. The *second phase* of pancreatitis involves the activation, chemoattraction, and sequestration of leukocytes and macrophages in the pancreas, resulting in an enhanced intrapancreatic inflammatory reaction. Neutrophil depletion induced by prior administration of an antineutrophil serum has been shown to reduce the severity of experimentally induced pancreatitis. There is also evidence to support the concept that neutrophils can activate trypsinogen. Thus, intrapancreatic acinar cell activation of trypsinogen could be a two-step process (i.e., an early neutrophil-independent and a later neutrophil-dependent phase). The *third phase* of pancreatitis is due to the effects of activated proteolytic enzymes and cytokines, released by the inflamed pancreas, on distant organs. Activated proteolytic enzymes, especially trypsin, not only digest pancreatic and peripancreatic tissues but also activate other enzymes such as elastase and phospholipase A₂. The active enzymes and cytokines then digest cellular membranes and cause proteolysis, edema, interstitial hemorrhage, vascular damage, coagulation necrosis, fat necrosis, and parenchymal cell necrosis. Cellular injury and death result in the liberation of bradykinin peptides, vasoactive substances, and histamine that can produce vasodilation, increased vascular permeability, and edema with profound effects on many organs. The systemic inflammatory response syndrome (SIRS) and acute respiratory distress syndrome (ARDS), as well as multiorgan failure, may occur as a result of this cascade of local and distant effects.

A number of genetic factors can increase the susceptibility and/or modify the severity of pancreatic injury in acute pancreatitis, recurrent pancreatitis, and chronic pancreatitis. All of the major genetic susceptibility factors center on the control of trypsin activity within the pancreatic acinar cell, in part because they were identified as candidate genes linked to intrapancreatic trypsin control. Five genetic variants have been identified as being associated with susceptibility to pancreatitis. The genes that have been identified include (1) cationic trypsinogen gene (*PRSS1*), (2) pancreatic secretory trypsin inhibitor (*SPINK1*), (3) the cystic fibrosis transmembrane conductance regulator gene (*CFTR*), (4) the chymotrypsin C gene (*CTRC*), and (5) the calcium-sensing receptor (*CASR*). Investigations of other genetic variants are currently under way, and new genes will be added to this list

in the future. Multiple medical, ethical, and psychological issues arise when these genes are discovered, and referral to genetic counselors is recommended.

APPROACH TO THE PATIENT:

Abdominal Pain

Abdominal pain is the major symptom of acute pancreatitis. Pain may vary from a mild discomfort to severe, constant, and incapacitating distress. Characteristically, the pain, which is steady and boring in character, is located in the epigastrium and periumbilical region, and may radiate to the back, chest, flanks, and lower abdomen. Nausea, vomiting, and abdominal distention due to gastric and intestinal hypomotility and chemical peritonitis are also frequent complaints.

Physical examination frequently reveals a distressed and anxious patient. Low-grade fever, tachycardia, and hypotension are fairly common. Shock is not unusual and may result from (1) hypovolemia secondary to exudation of blood and plasma proteins into the retroperitoneal space; (2) increased formation and release of kinin peptides, which cause vasodilation and increased vascular permeability; and (3) systemic effects of proteolytic and lipolytic enzymes released into the circulation. Jaundice occurs infrequently; when present, it usually is due to edema of the head of the pancreas with compression of the intrapancreatic portion of the common bile duct or passage of a biliary stone or sludge. Erythematous skin nodules due to subcutaneous fat necrosis may rarely occur. In 10–20% of patients, there are pulmonary findings, including basilar rales, atelectasis, and pleural effusion, the latter most frequently left sided. Abdominal tenderness and muscle rigidity are present to a variable degree, but compared with the intense pain, these signs may be less impressive. Bowel sounds are usually diminished or absent. An enlarged pancreas from acute fluid collection, walled off necrosis, or a pseudocyst may be palpable in the upper abdomen later in the course of the disease (i.e., 4–6 weeks). A faint blue discoloration around the umbilicus (Cullen's sign) may occur as the result of hemoperitoneum, and a blue-red-purple or green-brown discoloration of the flanks (Turner's sign) reflects tissue catabolism of hemoglobin from severe necrotizing pancreatitis with hemorrhage.

LABORATORY DATA

Serum amylase and lipase values threefold or more above normal virtually clinch the diagnosis if gut perforation, ischemia, and infarction are excluded. Serum lipase is the preferred test. However, it should be noted that there is no correlation between the severity of pancreatitis and the degree of serum lipase and amylase elevations. After 3–7 days, even with continuing evidence of pancreatitis, total serum amylase values tend to return toward normal. However, pancreatic isoamylase and lipase levels may remain elevated for 7–14 days. It should be recognized that amylase elevations in serum and urine occur in many conditions other than pancreatitis (see Chap. 370, Table 370-2). Importantly, patients with *acidemia* (arterial pH ≤ 7.32) may have spurious elevations in serum amylase. This finding explains why patients with diabetic ketoacidosis may have marked elevations in serum amylase without any other evidence of acute pancreatitis. Serum lipase activity increases in parallel with amylase activity and is more specific than amylase. A serum lipase measurement can be instrumental in differentiating a pancreatic or nonpancreatic cause for hyperamylasemia. *Leukocytosis* (15,000–20,000 leukocytes/ μ L) occurs frequently. Patients with more severe disease may show hemoconcentration with hematocrit values $>44\%$ and/or prerenal azotemia with a blood urea nitrogen (BUN) level >22 mg/dL resulting from loss of plasma into the retroperitoneal space and peritoneal cavity.

Hemoconcentration may be the harbinger of more severe disease (i.e., pancreatic necrosis), whereas azotemia is a significant risk factor for mortality. *Hyperglycemia* is common and is due to multiple factors, including decreased insulin release, increased glucagon release, and