end-stage liver disease can lead to worsening of hepatic decompensation, and HCV infection has recurred after transplantation in some of these recipients. Trials of hepatitis C immune globulin preparations to prevent recurrent hepatitis C after liver transplantation have not been successful. Similarly, a trial of a high-dose monoclonal antibody to the HCV E2 envelope glycoprotein delayed but did not prevent reappearance of viremia.

Although the current standard-of-care treatment of allograft hepatitis C is pegylated IFN and ribavirin, in a number of studies, the safety and efficacy of the addition of the approved HCV protease inhibitors telaprevir or boceprevir to pegylated IFN and ribavirin in genotype 1–infected patients with recurrent hepatitis C have been examined. Because of the profound inhibitory effects of the HCV protease inhibitors on the metabolism of the calcineurin inhibitors (increasing cyclosporine levels almost 5-fold and tacrolimus levels 70-fold), calcineurin inhibitor doses must be reduced to safe levels in these patients. In one multicenter study, treatment with a telaprevir- or boceprevir-based triple-drug regimen (with pegylated IFN and ribavirin) achieved rates of HCV clearance similar to those achieved in patients with chronic hepatitis C who had not undergone transplantation. Unfortunately, tolerability of these protease inhibitor–based regimens remains problematic in this population, particularly in persons with allograft cirrhosis, in whom the frequency of hepatic decompensation is increased. The approval of several new direct-acting antiviral (DAA) agents and of IFN-free DAA regimens against HCV will have a major impact on the management and outcome of both pretransplantation and posttransplantation HCV infection. Such therapeutic approaches (1) permit the clearance of viremia in a substantial proportion of decompensated cirrhotics, thereby preventing recurrent allograft infection and, possibly, even improving the clinical status of these patients, delaying or obviating the need for liver replacement; and (2) achieve sustained virologic responses in a much higher proportion of persons with allograft HCV infection, because of improvements in antiviral treatment efficacy and tolerability.

A small number of allograft recipients succumb to early HCV-associated liver injury, and a syndrome reminiscent of fibrosing cholestatic hepatitis (see above) has been observed rarely. Because patients with more episodes of rejection receive more immunosuppressive therapy, and because immunosuppressive therapy enhances HCV replication, patients with severe or multiple episodes of rejection are more likely to experience early recurrence of hepatitis C after transplantation. Both high viral levels and older donor age have been linked to recurrent HCV-induced liver disease and to earlier disease recurrence after transplantation.

Patients who undergo liver transplantation for end-stage alcoholic cirrhosis are at risk of resorting to drinking again after transplantation, a potential source of recurrent alcoholic liver injury. Currently, alcoholic liver disease is one of the more common indications for liver transplantation, accounting for 20–25% of all liver transplantation procedures, and most transplantation centers screen candidates carefully for predictors of continued abstinence. Recidivism is more likely in patients whose sobriety prior to transplantation was <6 months. For abstinent patients with alcoholic cirrhosis, liver transplantation can be undertaken successfully, with outcomes comparable to those for other categories of patients with chronic liver disease, when coordinated by a team approach that includes substance abuse counseling.

POSTTRANSPLANTATION QUALITY OF LIFE

Full rehabilitation is achieved in the majority of patients who survive the early postoperative months and escape chronic rejection or unmanageable infection. Psychosocial maladjustment interferes with medical compliance in a small number of patients, but most manage to adhere to immunosuppressive regimens, which must be continued indefinitely. In one study, 85% of patients who survived their transplant operations returned to gainful activities. In fact, some women have conceived and carried pregnancies to term after transplantation without demonstrable injury to their infants.

369 Diseases of the Gallbladder and Bile Ducts

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PHYSIOLOGY OF BILE PRODUCTION AND FLOW

BILE SECRETION AND COMPOSITION

Bile formed in the hepatic lobules is secreted into a complex network of canaliculi, small bile ductules, and larger bile ducts that run with lymphatics and branches of the portal vein and hepatic artery in portal tracts situated between hepatic lobules. These interlobular bile ducts coalesce to form larger septal bile ducts that join to form the right and left hepatic ducts, which in turn, unite to form the common hepatic duct. The common hepatic duct is joined by the cystic duct of the gallbladder to form the common bile duct (CBD), which enters the duodenum (often after joining the main pancreatic duct) through the ampulla of Vater.

Hepatic bile is an isotonic fluid with an electrolyte composition resembling blood plasma. The electrolyte composition of gallbladder bile differs from that of hepatic bile because most of the inorganic anions, chloride and bicarbonate, have been removed by reabsorption across the gallbladder epithelium. As a result of water reabsorption, total solute concentration of bile increases from 3–4 g/dL in hepatic bile to 10–15 g/dL in gallbladder bile.

Major solute components of bile by moles percent include bile acids (80%), lecithin and traces of other phospholipids (16%), and unesterified cholesterol (4.0%). In the lithogenic state, the cholesterol value can be as high as 8–10%. Other constituents include conjugated bilirubin; proteins (all immunoglobulins, albumin, metabolites of hormones, and other proteins metabolized in the liver); electrolytes; mucus; and, often, drugs and their metabolites.

The total daily basal secretion of hepatic bile is ~500–600 mL. Many substances taken up or synthesized by the hepatocyte are secreted into the bile canaliculi. The canalicular membrane forms microvilli and is associated with microfilaments of actin, microtubules, and other contractile elements. Prior to their secretion into the bile, many substances are taken up into the hepatocyte, while others, such as phospholipids, a portion of primary bile acids, and some cholesterol, are synthesized de novo in the hepatocyte. Three mechanisms are important in regulating bile flow: (1) active transport of bile acids from hepatocytes into the bile canaliculi, (2) active transport of other organic anions, and (3) cholangiocellular secretion. The last is a secretin-mediated and cyclic AMP–dependent mechanism that results in the secretion of a sodium- and bicarbonate-rich fluid into the bile ducts.

Active vectorial secretion of biliary constituents from the portal blood into the bile canaliculi is driven by a set of polarized transport systems at the basolateral (sinusoidal) and the canalicular apical plasma membrane domains of the hepatocyte. Two sinusoidal bile salt uptake systems have been cloned in humans, the Na^+/taurocholate cotransporter (NTCP, SLC10A1) and the organic anion–transporting proteins (OATPs), which also transport a large variety of non-bile salt organic anions. Several ATP-dependent canalicular transport systems, "export pumps," (ATP-binding cassette transport proteins, also known as ABC transporters) have been identified, the most important of which are: the bile salt export pump (BSEP, ABCB11); the anionic conjugate export pump (MRP2, ABCC2), which mediates the canalicular excretion of various amphiphilic conjugates formed by phase II conjugation (e.g., bilirubin mono- and diglucuronides and drugs); the multidrug export pump (MDR1, ABCB1) for hydrophobic cationic compounds; and the phospholipid export pump (MDR3, ABCB4). Two hemitransporters ABCG5/G8, functioning as a couple, constitute the canalicular cholesterol and phytosterol transporter. F1C1 (ATP8B1) is an aminophospholipid transferase ("flippase") essential for maintaining the lipid asymmetry of the canalicular membrane. The canalicular membrane also contains ATP-independent transport systems such as the Cl/HCO_3 anion exchanger isoform 2 (AE2, SLC4A2)