American Thoracic Society recommends that pulmonary infection due to NTM be diagnosed only when disease is clearly demonstrable—i.e., in an appropriate clinical and radiographic setting (nodules, bronchiectasis, cavities) and with repeated isolation of NTM from expectorated sputum or recovery of NTM from bronchoscopy or biopsy specimens. Given the large number of species of NTM and the importance of accurate diagnosis for the implementation of proper therapy, identification of these organisms is ideally taken to the species level.

The purified protein derivative (PPD) of tuberculin is delivered intradermally to evoke a memory T cell response to mycobacterial antigens. This test is variously referred to as the PPD test, the tuberculin skin test, and the Mantoux test, among other designations. Unfortunately, the cutaneous immune response to these tuberculosis-derived filtrate proteins does not differentiate well between infection with NTM and that with *M. tuberculosis*. Because intermediate reactions (~10 mm) to PPD in latent tuberculosis and nontuberculous mycobacterial infections can overlap significantly, the progressive decline in active tuberculosis in the United States means that NTM probably account for increasing proportions of PPD reactivity. In addition, bacille Calmette-Guérin (BCG) can cause some degree of cross-reactivity, posing problems of interpretation for patients who have received BCG vaccine. Assays to measure the elaboration of IFN-γ in response to the relatively tuberculosis-specific proteins ESAT6 and CFP10 form the basis for IFN-γ-release assays (IGRAs). These assays can be performed with whole blood or on membranes. It is important to note that *M. marinum, M. kansasii*, and *M. szulgai* also have ESAT6 and CFP10 and may cause false-positive reactions in IGRAs. Despite cross-reactivity with NTM, large PPD reactions (>15 mm) most commonly signify tuberculosis.

Isolation of NTM from blood specimens is clear evidence of disease. Whereas rapidly growing mycobacteria may proliferate in routine blood culture media, slow-growing NTM typically do not; thus it is imperative to suspect the diagnosis and to use the correct bottles for cultures. Isolation of NTM from a biopsy specimen constitutes strong evidence for infection, but cases of laboratory contamination do occur. Identification of organisms on stained sections of biopsy material confirms the authenticity of the culture. Certain NTM require lower incubation temperatures (*M. genavense*) or special additives (*M. haemophilum*) for growth. Some NTM (e.g., *M. tilburgii*) remain noncultivable but can be identified molecularly in clinical samples.

The radiographic appearance of nontuberculous mycobacterial disease in the lung depends on the underlying disease, the severity of the infection, and the imaging modality used. The advent and increase in the use of computed tomography (CT) has allowed the identification of characteristic changes that are highly consistent with nontuberculous mycobacterial infection, such as the "tree-in-bud" pattern of bronchiolar inflammation **(Fig. 204-2)**. Involvement of the lingual and right-middle lobes is commonly seen on chest CT but is difficult to appreciate on plain film. Severe bronchiectasis and cavity formation are common in more advanced disease. Isolation of NTM from respiratory samples can be confusing. *M. gordonae* is often recovered from respiratory samples but is not usually seen on smear and is almost never a pathogen. Patients with bronchiectasis occasionally have NTM recovered from sputum culture with a negative smear. The American Thoracic Society has developed guidelines for the diagnosis of infection with MAC, *M. abscessus*, and *M. kansasii*. A positive diagnosis requires the growth of NTM from two of three sputum samples, regardless of smear findings; a positive bronchoscopic alveolar sample, regardless of smear findings; or a pulmonary parenchyma biopsy sample with granulomatous inflammation or mycobacteria found on section and NTM found on culture. These guidelines probably apply to other NTM as well.

Although many laboratories use DNA probes to identify *M. tuberculosis*, MAC, *M. gordonae*, and *M. kansasii*, speciation of NTM helps determine the antimycobacterial therapy to be used. Only testing of MAC organisms for susceptibility to clarithromycin and of *M. kansasii* for susceptibility to rifampin is indicated; few data support other in vitro susceptibility tests, attractive though they appear. MAC isolates that have not been exposed to macrolides are almost always susceptible. NTM that have persisted beyond a course of antimicrobial therapy are often tested for antibiotic susceptibility, but the value and meaning of these tests are undetermined.

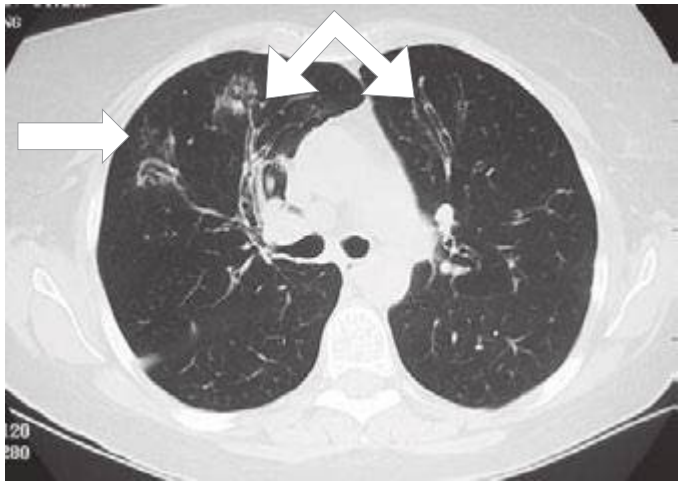

FIGURE 204-2 Chest computed tomography of a patient with pulmonary *Mycobacterium avium* complex infection. *Arrows* indicate the "tree-in-bud" pattern of bronchiolar inflammation (peripheral right lung) and bronchiectasis (central right and left lungs).

PREVENTION

Prophylaxis of MAC disease in patients infected with HIV is started when the CD4+ T lymphocyte count falls to <50/μL. Azithromycin (1200 mg weekly), clarithromycin (1000 mg daily), or rifabutin (300 mg daily) is effective. Macrolide prophylaxis in immunodeficient patients who are susceptible to NTM (e.g., those with defects in the IFN-γ/IL-12 axis) has not been prospectively validated but seems prudent.

TREATMENT NONTUBERCULOUS MYCOBACTERIA

NTM cause chronic infections that evolve relatively slowly over a period of weeks to years. Therefore, it is rarely necessary to initiate treatment on an emergent basis before the diagnosis is clear and the infecting species is known. Treatment of NTM is complex, often poorly tolerated, and potentially toxic. Just as in tuberculosis, inadequate single-drug therapy is almost always associated with the emergence of antimicrobial resistance and relapse.

MAC infection often requires multidrug therapy, the foundation of which is a macrolide (clarithromycin or azithromycin), ethambutol, and a rifamycin (rifampin or rifabutin). For disseminated nontuberculous mycobacterial disease in HIV-infected patients, the use of rifamycins poses special problems—i.e., rifamycin interactions with protease inhibitors. For pulmonary MAC disease, thrice-weekly administration of a macrolide, a rifamycin, and ethambutol has been successful. Therapy is prolonged, generally continuing for 12 months after culture conversion; typically, a course lasts for at least 18 months. Other drugs with activity against MAC organisms include IV and aerosolized aminoglycosides, fluoroquinolones, and clofazimine. In elderly patients, rifabutin can exert significant toxicity. However, with only modest efforts, most antimycobacterial regimens are well tolerated by most patients. Resection of cavitary lesions or severely bronchiectatic segments has been advocated for some patients, especially those with macrolide-resistant infections. The success of therapy for pulmonary MAC infections depends on whether disease is nodular or cavitary and on whether it is early or advanced, ranging from 20% to 80%.

M. kansasii lung disease is similar to tuberculosis in many ways and is also effectively treated with isoniazid (300 mg/d), rifampin (600 mg/d), and ethambutol (15 mg/kg per day). Other drugs with very high-level activity against *M. kansasii* include clarithromycin, fluoroquinolones, and aminoglycosides. Treatment should continue until