TABLE 169-5 PROPHYLACTIC REGIMENS COMMONLY USED TO DECREASE RISK OF INFECTION IN TRANSPLANT RECIPIENTS[a]

Risk Factor	Organism	Prophylactic Drug	Examination(s)[b]
Travel to or residence in area with known risk of endemic fungal infection	*Histoplasma*, *Blastomyces*, *Coccidioides*	Triazoles considered in context of clinical and laboratory assessment	Chest radiography, antigen testing, serology
Latent herpesviruses	HSV, VZV, CMV, EBV	Acyclovir after HSC transplantation to prevent HSV and VZV infection or reactivation; ganciclovir to prevent CMV infection, with possible effect on EBV/KSHV/HHV-6 infections in some settings	Serologic tests for HSV, VZV, CMV, HHV-6, EBV, KSHV; PCR
Latent fungi and parasites	*Pneumocystis jirovecii*, *Toxoplasma gondii*	Trimethoprim-sulfamethoxazole (or alternatives)	Serologic test for *Toxoplasma*
History of exposure to active or latent tuberculosis	*Mycobacterium tuberculosis*	Isoniazid in patients with recent seroconversion or positive chest imaging and/or no previous treatment	Chest imaging; TST and/or cell-based assay

[a]For information on latent infection with hepatitis B or C virus, see **Chap. 362**. [b]Serologic examination, tuberculin skin test, and interferon assays may be less reliable after transplantation.

Abbreviations: CMV, cytomegalovirus; EBV, Epstein-Barr virus; HHV-6, human herpesvirus type 6; HSC, hematopoietic stem cell; HSV, herpes simplex virus; KSHV, Kaposi's sarcoma–associated herpesvirus; PCR, polymerase chain reaction; TST, tuberculin skin test; VZV, varicella-zoster virus.

Prophylaxis with TMP-SMX for the first 4–6 months after transplantation decreases the incidence of early and middle-period infections (see below, Table 169-4, and **Table 169-5**).

Middle-Period Infections Because of continuing immunosuppression, kidney transplant recipients are predisposed to lung infections characteristic of those in patients with T cell deficiency (i.e., infections with intracellular bacteria, mycobacteria, nocardiae, fungi, viruses, and parasites). A high mortality rate associated with *Legionella pneumophila* infection **(Chap. 184)** led to the closing of renal transplant units in hospitals with endemic legionellosis.

About 50% of all renal transplant recipients presenting with fever 1–4 months after transplantation have evidence of CMV disease; CMV itself accounts for the fever in more than two-thirds of cases and thus is the predominant pathogen during this period. CMV infection **(Chap. 219)** may also present as arthralgias, myalgias, or organ-specific symptoms. During this period, this infection may represent primary disease (in the case of a seronegative recipient of a kidney from a seropositive donor) or may represent reactivation disease or superinfection. Patients may have atypical lymphocytosis. Unlike immunocompetent patients, however, they rarely have lymphadenopathy or splenomegaly. Therefore, clinical suspicion and laboratory confirmation are necessary for diagnosis. The clinical syndrome may be accompanied by bone marrow suppression (particularly leukopenia). CMV also causes glomerulopathy and is associated with an increased incidence of other opportunistic infections. Because of the frequency and severity of disease, a considerable effort has been made to prevent and treat CMV infection in renal transplant recipients. An immune globulin preparation enriched with antibodies to CMV was used by many centers in the past in an effort to protect the group at highest risk for severe infection (seronegative recipients of seropositive kidneys). However, with the development of effective oral antiviral agents, CMV immune globulin is no longer used. Ganciclovir (or valganciclovir) is beneficial for prophylaxis (when indicated) and for the treatment of serious CMV disease. The availability of valganciclovir has allowed most centers to move to oral prophylaxis for transplant recipients. Infection with the other herpesviruses may become evident within 6 months after transplantation or later. Early after transplantation, HSV may cause either oral or anogenital lesions that are usually responsive to acyclovir. Large ulcerating lesions in the anogenital area may lead to bladder and rectal dysfunction and may predispose the patient to bacterial infection. VZV may cause fatal disseminated infection in nonimmune kidney transplant recipients, but in immune patients reactivation zoster usually does not disseminate outside the dermatome; thus disseminated VZV infection is a less fearsome complication in kidney transplantation than in HSC transplantation. HHV-6 reactivation may take place and (although usually asymptomatic) may be associated with fever, rash, marrow suppression, or rare instances of renal impairment, hepatitis, colitis, or encephalitis.

EBV disease is more serious; it may present as an extranodal proliferation of B cells that invade the CNS, nasopharynx, liver, small bowel, heart, and other organs, including the transplanted kidney. The disease is diagnosed by the finding of a mass of proliferating EBV-positive B cells. The incidence of EBV-LPD is elevated among patients who acquire EBV infection from the donor and among patients given high doses of cyclosporine, tacrolimus, glucocorticoids, and anti–T cell antibodies. Disease may regress once immunocompetence is restored. KSHV infection can be transmitted with the donor kidney and result in development of Kaposi's sarcoma, although it more often represents reactivation of latent infection of the recipient. Kaposi's sarcoma often appears within 1 year after transplantation, although the time of onset ranges widely (1 month to ~20 years). Avoidance of immunosuppressive agents that inhibit calcineurin has been associated with less Kaposi's sarcoma, less EBV disease, and even less CMV replication. The use of rapamycin (sirolimus) has independently led to regression of Kaposi's sarcoma.

The papovaviruses BK virus and JC virus (polyomavirus hominis types 1 and 2) have been cultured from the urine of kidney transplant recipients (as they have from that of HSC transplant recipients) in the setting of profound immunosuppression. High levels of BK virus replication detected by PCR in urine and blood are predictive of pathology, especially in the setting of renal transplantation. JC virus may rarely cause similar disease in kidney transplantation. Urinary excretion of BK virus and BK viremia are associated with the development of ureteral strictures, polyomavirus-associated nephropathy (1–10% of renal transplant recipients), and (less commonly) generalized vasculopathy. Timely detection and early reduction of immunosuppression are critical and can reduce rates of graft loss related to polyomavirus-associated nephropathy from 90% to 10–30%. Therapeutic responses to IVIg, quinolones, leflunomide, and cidofovir have been reported, but the efficacy of these agents has not been substantiated through adequate clinical study. Most centers approach the problem by reducing immunosuppression in an effort to enhance host immunity and decrease viral titers. JC virus is associated with rare cases of progressive multifocal leukoencephalopathy. Adenoviruses may persist and cause hemorrhagic nephritis/cystitis with continued immunosuppression in these patients, but disseminated disease like that seen in HSC transplant recipients is much less common.

Kidney transplant recipients are also subject to infections with other intracellular organisms. These patients may develop pulmonary infections with *Mycobacterium*, *Aspergillus*, and *Mucor* species as well as infections with other pathogens in which the T cell/macrophage axis plays an important role. *L. monocytogenes* is a common cause of bacteremia ≥1 month after renal transplantation and should be seriously considered in renal transplant recipients presenting with fever and headache. Kidney transplant recipients may develop *Salmonella* bacteremia, which can lead to endovascular infections and require prolonged therapy. Pulmonary infections with *Pneumocystis* are common unless the patient is maintained on TMP-SMX prophylaxis.